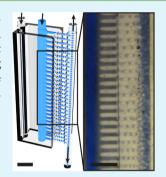
High-Throughput Generation of Emulsions and Microgels in Parallelized Microfluidic Drop-Makers Prepared by Rapid Prototyping

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Supporting Information

ABSTRACT: We describe the preparation of rapid prototyped parallelized microfluidic dropmaker devices. The manufacturing technique facilitates stacking of the drop-makers vertically on top of each other allowing for a reduced footprint and minimized dead-volume through efficient design of the distribution channels. We showcase the potential of the additive manufacturing technique for microfluidics and the performance of the parallelized device by producing large amounts of microgels with a diameter of ca. 500 μ m, a size that is inaccessible using traditional synthetic approaches.



KEYWORDS: microfluidics, microgel, upscaling, parallelization, 3D printing, rapid prototyping, additive manufacturing

1. INTRODUCTION

Monodisperse soft particles represent a versatile class of materials with manifold applications across disciplines. Small representatives of such particles in the nano- and micrometer regime can be produced using heterogeneous polymerization processes, such as emulsion and dispersion polymerization.^{1,2} Large uniform particles can only be prepared using microfluidic drop-makers, where the droplets act as templates for polymer particles with diameters ranging from micrometers to the millimeter scale.³⁻⁵ As the particles are generated one-by-one at a channel intersection inside of the drop-maker, the throughput of such devices is low and the yield of particles over time is poor. Several approaches have been pursued to parallelize dropmakers and increase the rate of droplet- and therefore particle generation. In *ladder*-type parallelization, the in- and outlets of a row of drop-makers are connected with distribution channels, extending the device geometry to a second layer for simple emulsions.⁶⁻⁸ This has been accomplished by carefully aligning and mating several layers of PDMS on top of each other. The rows of distribution channels can be connected in a third layer to further increase the amount of drop-makers per device. Using this technique, large amounts of double emulsions could be generated.⁶ By contrast, branched-channel devices rely on fluid distribution in the same layer as the channel intersection where the droplets are generated. Only the distribution of the dispersed phase is moved into the second layer, thus minimizing problems with alignment compared to ladder-type devices.9 Another way to minimize alignment processes and distribution channels is in circular drop-maker arrangements.^{10,11} Also combinations of these techniques have been applied with multiple circular arrangements connected with distribution channels, leading to the greatest number of paralellized drop-makers and highest throughput to date.¹²

All of these parallelization approaches exhibit a relatively large footprint, as all of the drop-makers are arranged in one layer. The two-dimensional layout entails large dead-volumes resulting from long distribution channels. The need for precise alignment of individual fluid-channel layers further obviates application of parallelized drop-makers in industrial processes.¹³ Stacking of drop-makers on top of each other while connecting the in- and outlets in the vertical direction would allow for a high density of microfluidic drop-makers per area and volume, while reducing the complexity of the device structure and minimizing dead volume. Current fabrication techniques for drop-maker parallelization, such as soft-lithography and micromachining, limit the structures to two-dimensional, obviating widespread use of parallelized devices.^{8,14}

Rapid prototyping based on additive fabrication principles enables the production of three-dimensional structures of nearly any desired shape or architecture.^{15–17} Different principles such as stereolithography, fused deposition molding, selective laser sintering or selective laser melting have been applied to generate microreactors for chemical synthesis.^{14,18–20} Direct rapid prototyping of microfluidic channel

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devices has recently been proposed, culminating in a modular system for plug-and-play microfludics.^{14,21–23} Direct rapid prototyping has become an important production technique because of its short design-to-device time, low cost, and full freedom of design.²⁴

Here we report the fabrication of a parallelized microfluidic flow-focusing device using digital light processing, a rapid prototyping technique that allows stacking of individual drop-makers on top of each other, minimizing the device footprint and distribution channel length. We showcase the applicability of our device by preparing monodisperse microgels with a diameter of ~500 μ m, which are inaccessible via standard emulsion or precipitation polymerization protocols.

2. RESULTS AND DISCUSSION

We start with designing the desired device architecture in a commercial CAD program. From the in- and outlets, fluid distribution channels extend into the device perpendicularly to the base area. The channel intersections at which droplets are generated are stacked vertically along the distribution channels, as depicted in Figure 1a, b (see also CAD files in the

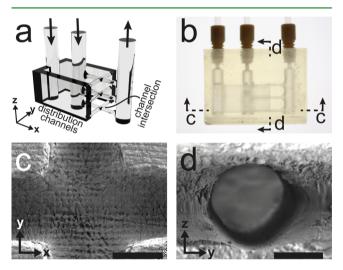


Figure 1. (a) Artistic rendering of the channel structure with inlets for continuous and dispersed phase and an outlet for the emulsion. (b) Photograph of prototyped device with tube connectors and dotted lines indicating the sectioning planes applied to image the SEM channel crossections in c and d. (c) Scanning electron micrograph of cross-section of the gold sputtered device allowing insight onto the channel contours showing voxels from micromirror array fabrication. The scale bar represents 500 μ m. (d) Close up on the channel intersection showing matching design and fabricated channel sizes. The scale bar represents 300 μ m.

Supporting Information). To achieve monodisperse droplet formation at all parallelized intersections, we need to pay special attention to pressure control inside of the device.²⁵ The device layout is predetermined by minimizing pressure variation along the different drop-maker intersections by designing the distribution channels with a much larger diameter than at the channel intersection, see Figure 1a. The channels leading to these intersections are circular with a radius of 375 μ m.

The 3D channel CAD architecture is fabricated by rapid prototyping, which builds up the structure layer-by-layer (Perfactory Mini-multilense from EnvisionTEC). Each layer contributes to the structure with a defined voxel-height, while the features within each layers are produced using a 180 W mercury vapor lamp, which is reflected off a micromirror array, illuminating 1400 \times 1050 pixels.^{26–29} We apply a hydrophobic methacrylate based photoresist (eShell600) with appropriate wetting properties and excellent solvent resistance for generating aqueous droplets in mineral oil. The methacrylate photoresist is exposed at 100 mW/dm² for 8 s in defined areas cross-linking the resist and hence producing the resulting structure at a resolution of $32 \times 32 \times 30 \ \mu m$ in *x-y-z*-directions. Depending on the aspect ratio, we can achieve a maximum resolution of 200 μ m for the channel width.^{27,28} The unexposed resist is removed subsequently using solvent treatment.²⁸ A cut through the device close to the channel intersection enables microscopic inspection of the indicated sectioning planes, as depicted in Figure 1b, c. The channels are accessible for the liquids and exhibit a voxel pattern in the x-y-plane, resulting from the micromirror lithography (see Figure 1c, d). The fabricated channel dimensions in fact match the design values (cf. CAD files in the Supporting Information).

We use Upchurch M-660 connections to guide the liquids to and away from the chip. Threads for access to the channel architecture are designed as part of the device manufacturing process. For the continuous phase we use a (50:50) mixture of mineral oil and heptane to obtain a suitable viscosity and we add Span 80 as a surfactant. As dispersed phase we use deionized water colored with blue ink (see video in the Supporting Information). Flow rates of the disperse and continuous phase as well as their ratios are known to influence the drop formation mechanism. We map out the droplet formation regimes for one of the drop-makers to explore the flow rates required to obtain monodisperse droplets. We obtain monodisperse droplets in the plug-flow and dripping regimes, see Figure 2a. Moving along Q_c^{100} for $Q_c/Q_d = 100$ we obtain a clear decrease in droplet size as we move from the plug-flow regime into the dripping regime and the aspect ratio (AR) approaches unity above 1100 μ L/min, see Figure 2b.

To verify that all three channel junctions produce droplets of the same size, we collected more than 100 droplets and performed image analysis, resulting in a standard deviation of 4.8% for the droplet diameter. Furthermore, the hexagonal arrangement of the collected droplets indicates monodispersity, see Figure 2c, d.

To showcase facile scalability of our concept, we prepared a device with 28 parallel cross-sections, as shown in Figure 3a. The previously described channel design with one inlet for the dispersed and continuous phase is extended by an additional outlet for the emulsion to compensate for varying pressure differences at individual junctions. The photograph image in Figure 3b demonstrates monodispersity of blue dyed water in oil droplets. The device easily produces droplet emulsions at rates of 3 L/h, suggesting utilization for industrial applications.

To demonstrate the applicability and superior performance of the prototyped parallel drop-maker device, we apply it to emulsify a microgel precursor together with a water-soluble photoinitiator (see the Supporting Information for details on synthesis). Subsequently, we perform a photochemical crosslinking reaction to generate monodisperse ~500 μ m microgels. Monodisperse microgels of such sizes are impossible to produce by standard heterogeneous polymerization techniques, as secondary processes give rise to bidisperse or polydisperse samples.² Here, a water-soluble six-armed star-shaped PEG precursor with a molecular weight of $M_n = 18$ kDa carrying acrylate groups at the terminus of each arm can be cross-linked photochemically. To cross-link the precursor inside of the

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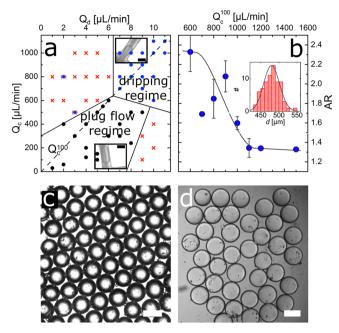


Figure 2. (a) Droplet formation map for the various conditions and different droplet formation regimes. The flow of the continuous phase Q_c versus the dispersed phase Q_{di} black circles: plug flow regime, blue circles: dripping regime, red crosses: no droplet formation. The blue circles with red parentheses represent conditions where droplets are formed sporadically. The inset micrographs display representative conditions at the channel intersection for the different regimes. (b) Droplet aspect ratio *AR* versus the flow of the continuous phase Q_c^{100} for $Q_c/Q_d = 100$. The inset shows the particle size distribution for the droplets in c. (c) Monodisperse water droplets in oil produced using the parallelized microfluidic device assemble in a hexagonal lattice indicating monodispersity. (d) Micrograph of microgels in aqueous medium. All scale bars represent 500 μ m.

microfluidically emulsified droplets the added radical photoinitiator is exposed to UV light with $\lambda_{max} = 380 \text{ nm.}^{30}$ The resulting microgels are large with dimensions similar to the droplets generated in the same device, see Figure 2d. This is the first example of high-throughput generation of microfluidically produced microgels. These unprecedented results substantiate the quality as well as versatility of our design and rapid prototyping technique.

3. CONCLUSION

The fabrication technique and parallelization concept presented here can be easily extended and adopted to other materials and other drop-maker geometries. The device is printed from a cross-linkable methacrylate, which exhibits excellent resistance toward polar and nonpolar solvents. The device geometry is compatible with techniques to further passivate the channel geometry using for example gas-phase deposition methods.⁵ The specific device architecture presented here facilitates the production of large droplets, which can act as reaction containers for chemical conversion, encapsulation or particle formation. In the future the challenge will lie in decreasing the channel diameters to generate smaller droplets and particles and achieve even higher drop-maker densities per chip. One potential way to conquer this demand could be the application of two-photon 3D lithography. However, also the highthroughput production of large diameter microgels, as presented here, is of special interest for several applications. These range from selective extraction and purification, to

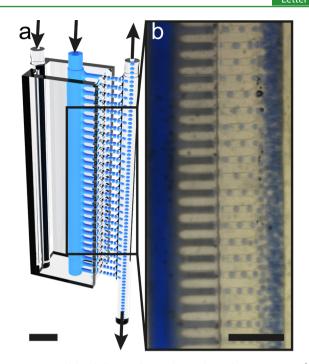


Figure 3. Parallelized drop-maker with 28 channel intersections. (a) Artistic rendering of an up-scaled parallel drop-maker with inlets for the dispersed and continuous phase and two outlets for the produced emulsion. (b) Micrograph of the running device producing blue dyed water droplets in oil. The scale bars represent 5 mm.

biomedical application in oral drug delivery or toxin uptake in the gastrointestinal passage. For these applications, it is crucial to control the microgel dispersity, while generating large amounts at the same time. The approach presented here will help to achieve these demanding requirements.

ASSOCIATED CONTENT

Supporting Information

Experimentals on the microfluidic emulsification and microgel preparation and video of the emulsification process in a parallelized triple drop-maker device. CAD files of the parallel triple and 28 dropmaker devices. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsami.5b03969.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Chern, C. S. Emulsion Polymerization Mechanisms and Kinetics. *Prog. Polym. Sci.* 2006, *31*, 443–486.

(2) Kawaguchi, S.; Ito, K. Dispersion Polymerization. *Adv. Polym. Sci.* 2005, 175, 299–328.

(3) Tumarkin, E.; Kumacheva, E. Microfluidic Generation of Microgels From Synthetic and Natural Polymers. *Chem. Soc. Rev.* **2009**, *38*, 2161–2168.

(4) Wang, W.; Zhang, M.-J.; Chu, L.-Y. Functional Polymeric Microparticles Engineered from Controllable Microfluidic Emulsions. *Acc. Chem. Res.* **2014**, *47*, 373–384.

(5) Kuehne, A. J. C.; Weitz, D. A. Highly Monodisperse Conjugated Polymer Particles Synthesized with Drop-based Microfluidics. *Chem. Commun.* **2011**, 47, 12379–12381.

(6) Romanowski, M. B.; Abate, A. R.; Rotem, A.; Holtze, C.; Weitz, D. A. High Throughput Production of Single Core Double Emulsions in a Parallelized Microfluidic Device. *Lab Chip* **2012**, *12*, 802–807.

(7) Kim, S.-H.; Kim, J. W.; Kim, D.-H.; Han, S.-H.; Weitz, D. A. Enhanced-throughput Production of Polymersomes Using a Parallelized Capillary Microfluidic Device. *Microfluid. Nanofluidics* **2012**, *14*, 509–514.

(8) Muluneh, M.; Issadore, D. Hybrid Soft-lithography/Laser Machined Microchips for the Parallel Generation of Droplets. *Lab Chip* **2013**, *13*, 4750–4754.

(9) Li, W.; Greener, J.; Voicu, D.; Kumacheva, E. Multiple Modular Microfluidic (M3) Reactors for the Synthesis of Polymer Particles. *Lab Chip* **2009**, *9*, 2715–2721.

(10) Nisisako, T.; Ando, T.; Hatsuzawa, T. High-volume Production of Single and Compound Emulsions in a Microfluidic Parallelization Arrangement Coupled with Coaxial Annular World-to-chip Interfaces. *Lab Chip* **2012**, *12*, 3426–3435.

(11) Bardin, D.; Kendall, M. R.; Dayton, P. A.; Lee, A. P. Parallel Generation of Uniform Fine Droplets at Hundreds of Kilohertz in a Flow-focusing Module. *Biomicrofluidics* **2013**, *7*, 034112.

(12) Conchouso, D.; Castro, D.; Khan, S. A.; Foulds, I. G. Three-Dimensional Parallelization of Microfluidic Droplet Generators for a Litre Per Hour Volume Production of Single Emulsions. *Lab Chip* **2014**, *14*, 3011–3020.

(13) Holtze, C. Large-scale Droplet Production in Microfluidic Devices - an Industrial Perspective. J. Phys. D: Appl. Phys. 2013, 46, 114008.

(14) Au, A. K.; Lee, W.; Folch, A. Mail-order Microfluidics: Evaluation of Stereolithography for the Production of Microfluidic Devices. *Lab Chip* **2014**, *14*, 1294–1301.

(15) King, P. H.; Jones, G.; Morgan, H.; de Planque, M. R. R.; Zauner, K.-P. Interdroplet Bilayer Arrays in Millifluidic Droplet Traps From 3D-printed Moulds. *Lab Chip* **2014**, *14*, 722–729.

(16) Comina, G.; Suska, A.; Filippini, D. PDMS Lab-on-a-chip Fabrication Using 3D Printed Templates. *Lab Chip* **2014**, *14*, 424– 430.

(17) McDonald, J. C.; Chabinyc, M. L.; Metallo, S. J.; Anderson, J. R.; Stroock, A. D.; Whitesides, G. M. Prototyping of Microfluidic Devices in Poly(dimethylsiloxane) Using Solid-Object Printing. *Anal. Chem.* **2002**, *74*, 1537–1545.

(18) Capel, A. J.; Edmondson, S.; Christie, S. D. R.; Goodridge, R. D.; Bibb, R. J.; Thurstans, M. Design and Additive Manufacture for Flow Chemistry. *Lab Chip* **2013**, *13*, 4583–90.

(19) Erkal, J. L.; Selimovic, A.; Gross, B. C.; Lockwood, S. Y.; Walton, E. L.; McNamara, S.; Martin, R. S.; Spence, D. M. 3D Printed Microfluidic Devices with Integrated Versatile and Reusable Electrodes. *Lab Chip* **2014**, *14*, 2023–2032.

(20) Symes, M. D.; Kitson, P. J.; Yan, J.; Richmond, C. J.; Cooper, G. J.; Bowman, R. W.; Vilbrandt, T.; Cronin, L. Integrated 3D-printed Reactionware for Chemical Synthesis and Analysis. *Nat. Chem.* **2012**, *4*, 349–354.

(21) Waldbaur, A.; Rapp, H.; Länge, K.; Rapp, B. E. Let There be Chip - Towards Rapid Prototyping of Microfluidic Devices: One Step Manufacturing Processes. *Anal. Methods* **2011**, *3*, 2681. (22) Kitson, P. J.; Rosnes, M. H.; Sans, V.; Dragone, V.; Cronin, L. Configurable 3D-Printed Millifluidic and Microfluidic 'Lab on a Chip' Reactionware Devices. *Lab Chip* **2012**, *12*, 3267–71.

(23) Donvito, L.; Galluccio, L.; Lombardo, A.; Morabito, G.; Nicolosi, A.; Reno, M. Experimental Validation of a Simple, Low-cost, T-junction Droplet Generator Fabricated Through 3D Printing. *J. Micromech. Microeng.* **2015**, *25*, 035013.

(24) Yuen, P. K.; Bliss, J. T.; Thompson, C. C.; Peterson, R. C. Multidimensional Modular Microfluidic System. *Lab Chip* 2009, 9, 3303–3305.

(25) Abrahamse, A. J.; van Lierop, R.; van der Sman, R. G. M.; van der Padt, A.; Boom, R. M. Analysis of Droplet Formation and Interactions During Cross-flow Membrane Emulsification. *J. Membr. Sci.* **2002**, *204*, 125–137.

(26) Melchels, F. P.; Feijen, J.; Grijpma, D. W. A Review on Stereolithography and its Applications in Biomedical Engineering. *Biomaterials* **2010**, *31*, 6121–6130.

(27) Femmer, T.; Kuehne, A. J. C.; Wessling, M. Print Your Own Membrane: Direct Rapid Prototyping of Polydimethylsiloxane. *Lab Chip* **2014**, *14*, 2610–2613.

(28) Femmer, T.; Kuehne, A. J. C.; Torres-Rendon, J.; Walther, A.; Wessling, M. Print Your Membrane: Rapid Prototyping of Complex 3D-PDMS Membranes via a Sacrificial Resist. *J. Membr. Sci.* **2015**, *478*, 12–18.

(29) Torres-Rendon, J. G.; Femmer, T.; De Laporte, L.; Tigges, T.; Rahimi, K.; Gremse, F.; Lederle, W.; Ifuku, S.; Wessling, M.; Hardy, J. G.; Walther, A. Bioactive Gyroid Scaffolds Formed by Sacrificial Templating of Nanocellulose and Nanochitin Hydrogels as Instructive Platforms for Biomimetic Tissue Engineering. *Adv. Mater.* **2015**, *27*, 2989–2995.

(30) Seiffert, S.; Weitz, D. A. Controlled Fabrication of Polymer Microgels by Polymer-analogous Gelation in Droplet Microfluidics. *Soft Matter* **2010**, *6*, 3184–3190.